

Novel antiretroviral therapeutic strategies for HIV

Rita F Cunha¹, Sandra Simões^{2*}, Andreia Ascenso^{2*} and Quirina Costa³

¹Faculty of Pharmacy, University of Lisbon, Portugal

²Department of Pharmaceutical Technology, Portugal

³Department of Microbiology and immunology, Portugal

Abstract

When the first cases of HIV infection first appeared in the 1980s, AIDS was a deadly disease without any therapeutic alternatives. Currently, there is still no cure for most cases mainly due to the multiple tissues that act as a reservoir for this virus besides the high viral mutagenesis that leads to in antiretroviral drug resistance. Throughout the years, multiple drugs with specific mechanisms of action on distinct targets have been approved. In this review, the most recent phase III clinical studies and other research therapies as advanced transdermal antiretroviral nanodelivery systems will be here discussed in the HIV context.

Although the combined antiretroviral therapy is effective in reducing viral loading to undetectable levels, it also presents some disadvantages, such as usual side effects, high frequency of administration, and the possibility of drug resistance. In this context, several new drugs and delivery systems (including novel targets) and vaccines have been tested in pre-clinical and clinical trials. Regarding drug delivery, an attempt to change the route of administration of some conventional antiretrovirals has proven to be successful and surpassed some issues related to patient compliance. In particular, nanotechnology has brought a new approach to overcoming certain obstacles of formulation design including drug solubility and biodistribution. Overall, the encapsulation of antiretroviral drugs into nanosystems has shown improved drug release and pharmacokinetic profile.

Keywords: HIV; clinical trials; novel antiretrovirals; vaccines; advanced transdermal nanodelivery systems

Abbreviations: AIDS: Acquired Immunodeficiency Syndrome; ABC: Abacavir; ADCC: Anti-body dependent cellular cytotoxicity ; ART: antiretroviral therapy; ARV: antiretroviral ; ATZ: Atazanavir; AZT: Zidovudine ; CA: Capsid protein; CAP: Cellulose Acetate-Phthalate; CNS: Central Nervous System; CR: cabotegravir ; c-ART: combination antiretroviral therapy; cDNA: complementary DNA; CD4 bs: CD4 binding site; CHR: C-terminal heptad repeat; DLV: Delaviridine; DTG: Dolutegravir; EFV: Efavirenz; EMA: European Medicines Agency; Env: Envelope Glycoprotein; ETR: Etravirine; FDA: Food and Drug Administration; FDC: Fixed dose combination; FI: Fusion Inhibitors; FP: Fusion peptide; FPPR: Fusion peptide proximal region; Gag: group specific antigen; GALT: gut-associated lymphatic tissue; GFR: Glomerular filtration rate; HeLa: human cervical cells; HIV: human immunodeficiency virus; IN: integrase; II: Integrase Inhibitors; INSTI: Integrase Strand Transfer Inhibitors; LA: Long acting; LC.MS/MS: liquid chromatography/electrospray ionization mass spectrometry; 3TC: Lamivudine; LTR: long-term repeat; MA: Matrix protein; mAb: Monoclonal antibodies; MPER: Membrane proximal external region; MVC: Maraviroc; nAb: Neutralizing antibodies; NC: Nucleocapsid; ND: nanodiamond ; NHR: N-terminal heptad repeat; NRTI: Nucleotide/nucleoside Reverse Transcriptase Inhibitors; NSI: Non-syncytium inducing; OBT: Optimized background therapy; PGLA: poly (lactide co-glycolide); NFV: Nelfinavir; NNRTI: Non-nucleoside reverse transcriptase Inhibitors; NRC: non-randomized cohort; NVP: Nevirapine; PBD: Pocket-binding domain; PBMC: Peripheral blood mononuclear cells ; PI: Protease Inhibitors; PIB: Polyisobutylene; Pol: Polymerase; PETIM: poly (propyl ether imine); PR: Protease; pM: picomolar; PSA: Pressure sensitive adhesives; RAL: Raltegravir; RC: Randomized cohort; RPV: Rilpivirine; RT: reverse transcriptase; SI: syncytium inducing; 6 HB: Six-helix bundle; SIV: Simian immunodeficiency viruses; SQV: Saquinavir; TAF: Tenofovir Alafenamide; TAR: Trans-activation response element; TCR: T-cell receptor; TDDS: Transdermal Drug Delivery System; TMC114: Darunavir; TMD: Transmembrane domain; TNF: Tenofovir disoproxil fumarate; TRM: tryptophan rich motif

Introduction

The human immunodeficiency virus (HIV) is still a very prominent disease worldwide. Acquired Immunodeficiency Syndrome (AIDS) can now be considered a chronic infection since patients are living longer due to the several options of antiretroviral therapy [1]. The number of new cases has decreased from 3.3 million in 2002 to 1.6 million in 2012, nevertheless, an estimated 37.9 million people are living with HIV and 1.7 million

are newly infected [1,2].

The HIV genome is formed by two identical single chain RNA molecules, which are confined to the core of the virus particle. The virus particle produces an enzyme, known as reverse transcriptase (RT), responsible for the transcription of the viral RNA into the pro-viral (double-chained) DNA, which can be integrated into the human genome due to the function of another viral protein, the integrase (IN).

The viral genome is delimited at both ends by long-term repeat sequences (LTR). The viral gene transcription is followed by the group-specific antigen (gag) gene, which codes for the outer core matrix protein (MA/p17), the capsid protein (CA/p24), the nucleocapsid, and the nucleic-acid stabilizing protein. The gag gene is followed by the polymerase (pol) gene, which is responsible for the production of RT, the RNase H, and viral integrase. The envelope (env) gene codes for two membrane proteins gp 140 (transmembrane protein, TM) and gp120 (surface glycoprotein SU) which is composed of five variable regions (loops V1 to V5) and five preserved regions (C1 to C5) [3]. Initially, Env binds to the host protein CD4 of T helper cells, macrophages, astrocytes, and dendritic cells by interacting with the CD4 binding site (CD4 bs) through its gp120 subunit. In turn, this interaction causes conformational changes in the variable regions. The conformational change in V3 and the formation of β -sheet induce a conformational change that exposes a new site for the gp120 subunit to bind the co-receptor [3,4]. The co-receptor binding causes a change in the gp 41 subunit by disclosing a gp 41 fusion protein (one for each gp41 in the trimer), which will connect to the target cell membrane due to its extremely hydrophobic nature, thereby enabling the fusion of both viral envelope and host cell membrane [4]. Following the fusion of both virion and host cell, the viral capsid is then absorbed by an endosome. Inside the phagocyte, H⁺ ions are released, causing a decrease in pH value, and inducing the delivery of the capsid into the cytoplasm. The viral RT will convert the viral single-stranded RNA into complementary DNA (cDNA), forming a pre-integration complex, which can be integrated into the host cell genome, while the RNA strand is destroyed by RNase H [3]. The cDNA is transported through the cytoplasm and into the host cell nucleus via the nucleopores, with the intervention of the Vpr protein. This process is fundamentally mediated by the integrase protein (IN) starting by removing nucleotides from the 3' ends of the proviral DNA, and then, proceeding to catalyze a nucleophilic attack to the phosphodiester bonds of the DNA chains, thus forming a covalent bond between viral and host DNA. This is the essential step in viral replication that allows the viral proteins to be produced along with those necessary for cellular function [6,7]. Once the integration is completed, several viral proteins can be produced through transcription using the RNA polymerase II of the host cell. Transcription factors bind to the LTR regions and, after a chain of chemical reactions, the regulatory proteins such as Tat, Rev, and Nef are produced. Tat will activate transcription by connecting its trans-activation response element (TAR) element to the LTR region [6,7]. The Gag-Pol protein resulting from the translation of mRNA is transported and becomes embedded into the plasma membrane of the host cell [8]. Meanwhile, all the other structural viral proteins are produced and ensembled together in the plasma membrane, forming a multiprotein structure. This leads to the activation of the viral protease (PR), which will allow the release of all structural proteins, such as MA, CA and NC, as well as their reorganization into mature virions [6,7].

The HIV virions can be classified as R5 viruses if they have tropism for the CCR5 co-receptor, or as X4 viruses, if they interact with the CXCR4 co-receptor. It might be possible that a viral particle uses both co-receptors to infect the target cell, and in this case, it would be classified as an R5X4 virus. Generally, R5 viruses are more abundant at the early stages of infection, while X4 or R5X4 viral strains predominate at later stages [4,5]. The most common change in tropism is the alteration from R5 to X4, usually followed by CD4⁺ cell counts dropping, but the reverse can also occur.

The period between the infection of the first host cell and the

detection of the virus in the blood is called the eclipse phase, and usually lasts from 7 to 21 days. After the infection of the first cell, the virus continues to replicate in the mucosa, submucosa, and adjacent lymphatic tissue. The replication concentrates in the gut-associated lymphatic tissue (GALT) quite early [9,10]. Then, it follows an exponential rise of the viral loading in which the CD4⁺ cell counting rapidly decreases. This phase is characterized by flu-like and non-specific clinical signs that usually last between 7 to 10 days. After a few weeks, the immune system can generate a response [6,11]. The cellular immune response starts with the activation of CD8⁺ cytotoxic lymphocytes. Their T-cell receptor (TCR) will bind to viral proteins, which are in turn connected to the antigen-presenting molecule (MHC I) to eliminate the infected cells [6]. Generally, after 3 to 5 weeks, a humoral response starts to produce specific neutralizing antibodies that will destroy the virions via phagocytosis. The convergence of both types of immune responses leads to a decrease in viremia and a new rise in CD4⁺ cell count. The period, in which there is an infection but without antibodies, is called the "serological window period" [11]. Even though this is an asymptomatic phase, and the viral loading is somewhat controlled, there is still a loss of immune cells since the virus continues to replicate in the lymphatic tissue (its reservoir), destroying its structure. Then, the viral loading becomes higher as the CD4⁺ cell count diminishes, leading to the beginning of the AIDS stage. In this stage, the patients are more susceptible to opportunistic infections such as *Mycobacterium tuberculosis*, *Cytomegalovirus*, *Herpes Zoster*, and Kaposi Sarcoma, among many others [11].

Several aspects make it difficult to eradicate the virus once a patient is infected. One of them is related to the absence of proofreading activity in the viral RT, causing a great number of mutations and genetic diversity in the HIV genome. The other one is concerned with the ability of the virus to infect resting memory or naïve cells, leading to a latent viral state [6,11]. The problem with viral latency is that it can occur even after patients have undergone antiretroviral therapy reducing viremia to an undetectable level [6].

Therapeutic Targets vs Commercial Antiretrovirals

Based on the replication cycle of HIV, several therapeutic targets and antiretroviral drugs have been developed over the years. Nowadays, any initial therapy regimen must have at least three different drugs with distinct therapeutic targets. In general, antiretrovirals can be classified into eight major types, according to their mechanism of action [12,13]:

- 2.1. Nucleotide/Nucleoside Reverse Transcriptase Inhibitors (NRTI)
- 2.2. Non- Nucleotide Reverse Transcriptase Inhibitors (NNRTI)
- 2.3. Integrase Inhibitors (II)
- 2.4. Protease Inhibitors (PI)
- 2.5. Fusion Inhibitors (FI)
- 2.6. Pharmacokinetic Enhancers (PE)
- 2.7. CCR5 Antagonist

Numerous examples of these antiretroviral drugs are listed in (Table 1).

Nucleotide/nucleoside Reverse Transcriptase Inhibitors (NRTI)

The first drugs to be developed were the *Nucleotide/Nucleoside Reverse Transcriptase Inhibitors* (NRTI). Their structure is very similar to the viral nucleosides, except absence of a hydroxyl group in the 3' position of their deoxyribose sugar, thereby preventing a phosphodiester bond between the NRTIs and the next 5' nucleosides. As a result, the nucleoside chain is interrupted and

Table 1. Classification of commercial antiretroviral drugs for HIV and therapeutic advantages and disadvantages.

Class of Antiretroviral Drugs	Therapeutic Target	Approved Drugs	Advantages	Disadvantages	References
Nucleotide/Nucleoside Reverse Transcriptase Inhibitors	Reverse Transcriptase	Abacavir (ABC) Ziagen® Tenofovir disoproxil fumarate (TNF) Viread® Lamivudine (3TC) Epivir® Emtricitabine (FTC) Emtriva® Zidovudine (AZT) Retrovir®	Long intracellular half-life period; High oral bioavailability; Few interactions; No problems with administration	Highly prone to resistance Adverse effects: myelosuppression neuropathy pancreatitis, nausea, vomiting fatigue, anemia lactic acid accumulation	[15] [13]
Non- Nucleotide Reverse Transcriptase Inhibitors	Reverse Transcriptase (viral replication)	Efavirenz (EFV) Sustiva® Nevirapine (NVP) Viramune® Delavirdine (DLV) Rescriptor® Etravirine (ETR) Intelence® Ralpivirine (RPV) Edurant®	More selective than NRTIs; Cheaper to produce; Single-tablet regimens	Very prone to resistance. Rash, nausea, vomiting, fatigue, mood swings, depression, jaundice, conjunctivitis and respiratory issues	[14] [16] [17]
Integrase Inhibitors	Viral Integrase (DNA integration)	Raltegravir (RAL) Isentress®	Very selective drugs, they interact with two components of viral replication.	Hypersensitivity reactions, rash, jaundice, dark-colored urine, nausea, vomiting, fatigue, blisters in the mouth and skin, diarrhea and loss of appetite.	[13] [14] [16]
		Dolutegravir (DTG) Tivicay®			
Protease Inhibitors	Viral protease (protein synthesis)	Ritonavir (RTV) Norvir® Nelfinavir (NFV) Viracept® Atazanavir (ATZ) Reyataz® Darunavir (TMC114) Prezista® Saquinavir (SQV) Invirase® Fortovase®	Active against HIV-1 and HIV-2	High prevalence of resistance; Arrhythmia, heartburn, fatigue, jaundice, dizziness, abdominal pain, mouth sores and urinary tract issues	[14] [16] [18] [13]
CCR5 antagonist	CCR5 co-receptor	Maraviroc Celsentri®	Effective in cases of resistance to conventional therapy regimens	Only effective in R5 viruses; Nausea, Diarrhea, Fatigue, headache	[19] [20]
Post-attachment inhibitors	CD4+ cells	Ibalizumab Trogarzo®	Pharmacokinetics allows for a weekly administration; Does not cause CD4 depletion; No evidence of resistance	Immune reconstitution inflammatory syndrome	[14] [21]
Pharmacokinetic Enhancers	CYP3A subfamily	Cobicistat Tybost®	More selective than ritonavir; Less drug-drug interactions	Might cause raises in serum creatinine; Side effects in the gastrointestinal tract	[22] [23] [14]
Fusion Inhibitors	gp41 subunit	Enfuvirtide Fuzeon®	Decreases viral load in a c-ART regimen; Increases CD4 cell counts; Low toxicity; High specificity;	Short half-life; Low threshold for drug resistance; Can cause reaction in its injection-site, nausea and fatigue; High cost; Inconvenient route of administration	[15] [24] [25] [26] [27]

the proviral DNA is not formed. These drugs are formulated and delivered as prodrugs, which need to be phosphorylated to become active [12,15]. NRTIs are classified as competitive inhibitors of the RT, as they bind to the enzyme active site being integrated into the viral DNA chain [17].

Regarding pharmacokinetics, this class of drugs is known to have a long intracellular half-life period, good oral bioavailability, and no restrictions of administration, and improbable interactions with other drugs [14]. On the other hand, few side effects have been documented such as myelosuppression, pancreatitis, and neuropathy. However, the resistance mechanisms have been noticed as well [12,14].

The first NRTI approved was Zidovudine (AZT) in 1987 and the last one was Emtricitabine in 2003 [27].

Non-nucleoside Reverse Transcriptase Inhibitors (NNRTIs)

The *Non-nucleoside Reverse Transcriptase Inhibitors* (NNRTIs) bind directly to the RT in a non-competitive inhibition process [13]. NNRTIs bind to a specific pocket of the viral RT, away from the active site, thereby inducing a conformational change that inhibits the enzymatic activity [17]. NNRTIs do not inhibit the RT of any other retroviruses, nor one of HIV-2 [15]. NNRTIs are quite cheap to produce and allow the single tablet regimens [16]. These drugs can cause rashes, nausea, vomiting, fatigue, mood swings depression, jaundice, conjunctivitis, and respiratory issues [13]. Just like the NRTIs, NNRTIs are very prone to inducing resistance [15].

The first NNRTI to be approved was Nevirapine (NVP) in 1996, the same year when combined antiretroviral therapy (c-ART) was first introduced [16]. The most recent drug to be approved was Doravirine in November 2018 [28].

Integrase Inhibitors (IIs)

In the last decade, the viral integrase has been successfully used as a therapeutic target for HIV through a unique biochemical mechanism. The integrase inhibitors designed until now are *strand transfer reaction inhibitors* (INSTIs). These inhibitors are quite selective drugs since they interact with the co-factors (metallic cations) at the active site of this enzyme, and then, with the complex formed by viral DNA and integrase. The pharmacophore binds to the metallic cations whereas a lipophilic group interacts with the viral DNA-integrase complex [15].

INSTIs can cause hypersensitivity reactions, rash, jaundice, dark-colored urine, nausea, vomiting, fatigue, blisters in the mouth and skin, diarrhea, and loss of appetite [13].

Raltegravir (RAL) was the first integrase inhibitor approved in 2007. The most recent approval was Elvitegravir in 2014 [16]. However, its marketing authorization in the European Union was withdrawn in 2016 for commercial reasons [29].

Protease Inhibitors (PIs)

Protease inhibitors bind to this enzyme in a competitive manner as its natural substrate. The inhibition of this enzymatic activity leads to immature virions and less viral spreading [15,17].

The main inconvenience of these antiretroviral drugs is that the protease gene is very prone to mutation, which can easily lead to resistance [15].

PIs are known to cause arrhythmia, heartburn, fatigue, jaundice, dizziness, abdominal pain, mouth sores, kidney stones, and dark-colored urine [13].

Saquinavir (SQV) was the first PI to be approved in 1995 by the Food and Drug Administration (FDA) and in 1996 by the European Medicines Agency (EMA) [28,30]. The most recent approval for this drug category was Darunavir in 2006 by the FDA and in 2007 by the EMA [27,28].

Fusion inhibitors

Enfuvirtide (Fuzeon[®]) was the first drug approved as a fusion inhibitor (gp41). The gp41 component can be sub-divided into the N-terminal heptad repeat (NRH)- binding domain which is analogous to Fuzeon[®], and a pocket-binding domain (PBD), with hydrophobic pockets that bind to the NHR exterior [23]. This drug acts by inhibiting the connection between NHR (or HR-1) and the C-terminal heptad repeat (CHR or HR-2), thereby preventing the formation of the 6 helix-bundle (6HB) or “core”, and ultimately, the membrane fusion with the viral envelope [23].

Enfuvirtide was approved in 2003 as a treatment option for patients infected with HIV-1 who were in an advanced stage of disease progression and were resistant to other antiretroviral drug classes [25]. It is administered by subcutaneous injection and the standard regimen is 90 mg twice a day [26]. The most common side effects are reactions near the injection site, nausea, diarrhea, fatigue, and eosinophilia [26].

The TORO clinical trials demonstrated few benefits in the administration of enfuvirtide in cases where patients have shown resistance to therapy with NRTIs [26]. The viremia and CD4 cell counts were compared in patients taking both enfuvirtide and an optimized conventional ART regimen versus only the conventional ART regimen. After 48 weeks of treatment, 30% of the enfuvirtide patients had their viremia reduced to <400 copies/ml versus 12% in the conventional therapy arm. At a 95 week follow-up, 17.5% of patients taking enfuvirtide had their viral loading under 50 copies/ml [26].

Despite being a promising therapeutic option, it presents certain disadvantages. Firstly, it has a very short half-life, which leads to an inconvenient posology and administration route, and consequent low patient compliance. Secondly, it has a very low antiretroviral activity and only active against HIV-1. Finally, this drug has a very low threshold for resistance [24,31]. The resistance occurs mainly due to mutations in the HR-1 region of gp41, even though mutations in HR-2 are also possible [14,26].

In order to surpass these issues, several enfuvirtide-based lipopeptides have been synthesized over the years including LP40, LP46, LP52, and LP80 as second-generation fusion inhibitors. Although LP46 presented higher antiviral activity than enfuvirtide, its phase II clinical trial was interrupted due to reports of serious side effects [32]. LP52 was designed based on LP46 and LP40 exhibiting a higher threshold for drug resistance and capable of inhibiting resistant strains. Additionally, it has a longer half-life and higher antiviral activity with low IC₅₀ values (pM range) [32]. In a recent study [33], several new lipopeptides were developed using fatty acids with different lengths, for example, LP80 which was found to be a quite potent fusion inhibitor against resistant strains. It showed similar cytotoxicity compared to LP52 and enfuvirtide in different cell lines, such as TZM-bl cells, HEK293T cells, and MT-4 cells. Other promising results were obtained in pharmacokinetic studies in healthy rhesus macaques [33]. When administered at 3mg/kg to rhesus macaques chronically infected with simian-human immunodeficiency virus (SHIV), once a day for two weeks, viral loading was reduced to below the detection limit after 4 days of treatment in 3 out of 5 monkeys [33].

Pharmacokinetic enhancers

Cobicistat was approved in 2012 as a pharmacokinetic booster in co-administration with other antiretrovirals. Contrarily to ritonavir, it has no specific antiretroviral activity. However, it is substantially more selective than ritonavir, considering that it inhibits only the CYP3A isoenzyme subfamily, leading to fewer drug interactions [21,22]. By inhibiting the CYP3A isoenzymes, cobicistat allows

an increase in the plasma concentrations of other antiretrovirals, such as PIs and NNRTIs, thereby enabling higher intervals in administration with a lesser pill burden and improved adherence to therapy [22].

Some clinical trials showed an increase of serum creatinine levels and a decrease in glomerular filtration rate (GFR) in patients taking cobicistat. Nevertheless, in a phase I study, these abnormalities were explained due to a decline in the activity of certain transporters, such as SLC22A2, which are essential for the elimination of creatinine. This means that cobicistat causes higher serum creatinine levels, thereby altering the GFR estimation calculated using the Cockcroft-Gault formula. Thus, it does not really alter GF [21,22]. It is now known that this drug restrains some cation renal transporters. However, no dosage adjustments are required for patients with defective liver or kidney function [21].

Regarding CYP3A4 inducers, such as carbamazepine and rifabutin, cobicistat is not as effective when administered simultaneously with these drugs, since it is mostly metabolized by CYP3A4 and CYP2D6 [21].

CCR5-antagonist

CCR5- antagonists were the first class of antiretroviral drugs to target host cells and not virions [18]. In fact, CCR5 is the most common co-receptor present in the majority of target cells in the early stage, which allows it to be a very promising therapeutic target for that stage [18].

The CCR5-antagonist Maraviroc (MVC) was approved by both FDA and EMA in 2007 [28]. MVC binds to the hydrophobic pocket of the co-receptor inducing a conformational change that avoids its recognition by viral gp120, thereby limiting virion entry into peripheral blood mononuclear cells (PBMC) [12,34]. MVC is mostly metabolized in the liver by CYP450 enzymes, such as CYP3A4 and CYP3A5, which is why it should be administered cautiously with other CYP450 inducing or inhibiting drugs, such as PIs [34]. The most common adverse effects are nausea, diarrhea, fatigue, and headaches [19]. Furthermore, it is known some mechanisms of resistance related to a higher affinity of gp120 towards CCR5 [34].

At least, some examples of combination antiretroviral therapy (c-ART) are listed in (Table 2). c-ART was first implemented in 1996, and since then, some disadvantages were found as the interference of PIs and NNRTIs with the metabolization process of other common drugs [22]. Ritonavir started to be progressively co-administered with other first-generation PIs, such as lopinavir or saquinavir, and afterward, with second-generation PIs, including atazanavir and darunavir [22]. As ritonavir could constrain the metabolization process, higher concentrations of other antiretrovirals and larger intervals in their administration were possible to achieve.

New Drugs and Vaccines – Overview of Phase III Clinical Trials

Nowadays the wide range of therapeutic options and c-ART regimens have allowed that AIDS become a chronic disease with lower mortality [41]. Although c-ART is the best therapeutic strategy until now, it is far away to be the ideal solution considering that patients are forced to life-long medication, which presents a considerable risk of resistance emergence and rebound in viral loading. Moreover, these patients also suffer from numerous side effects [42,43].

When discussing a cure for the infection of the HIV virus, it is important to establish the difference between a “sterilizing cure” and a “functional cure”. A “sterilizing cure” consists of the elimination of both actively infected cells and the virus latent reservoir, whereas a “functional cure” implies a prolonged suppression of viremia to an undetectable level and sustaining a normal CD4+ cell count [41].

There are currently several new drugs and vaccines being tested in pre-clinical and clinical trials for HIV treatment or prophylaxis. Hereby, the most recent clinical studies on phase III will be further discussed.

Post-attachment inhibitors: Ibalizumab

Ibalizumab was granted a “breakthrough therapy status” in 2015 and it was approved in March 2018 by the FDA [44]. Its approval by the EMA is still pending at this time, even though this agency has issued a positive opinion, recommending the granting of a marketing authorization for this drug in 2019 [45].

As an IG4 monoclonal antibody (IG4 mAb), ibalizumab connects to domain 2 of the CD4 receptor and leads to conformational changes which avert the gp 120 – co-receptor interactions, thereby preventing the viral entry [47]. This innovative drug presents a very low ADCC (antibody dependent cellular cytotoxicity), and hence a quite low rapport towards cytotoxic immune responses. Therefore, CD4+ cells will not be destroyed through cytotoxic pathways [47].

Ibalizumab is used in combination with other drugs for the treatment of non-naïve patients, who have already been unsuccessfully treated with other classes of antiretrovirals due to drug resistance. It is the first drug to target the CD4 receptor, and therefore, the first one to be considered a post-attachment inhibitor [44,46]. The FDA recommends the subcutaneous administration of 2000 mg loading dose, and subsequently, 800 mg maintenance dose every 2 weeks [44]. The half-life of this drug is estimated to range between 3 and 3.5 days [20,47]. Ibalizumab is generally well tolerated, with some adverse effects such as rash, headaches, nausea, and depression, reported by a low percentage of patients [47].

Table 2. Examples of combination antiretroviral therapy for HIV.

Approved Drugs	Active Substances	References
Genvoya*	150 mg elvitegravir/ 150 mg cobicistat/ 200 mg emtricitabine/ 10 mg tenofovir	[36]
Atripla*	600 mg efavirenz/ 200 mg emtricitabine/ 245 mg tenofovir-DF	[37]
Rezolsta*	800 mg darunavir/ 150 mg cobicistat	[38]
Triumeq*	50 mg dolutegravir/ 600 mg abacavir/ 300 mg lamivudine	[39]
Evotaz*	300 mg atazanavir / 150 mg cobicistat	[40]
Descovy*	200 mg emtricitabine / 10 mg tenofovir alafenamide 200 mg emtricitabine / 25 mg tenofovir alafenamide	[41]

In summary, ibalizumab presents as a very promising and safe therapeutic option for patients suffering from resistance to the recommended ARV regimens, and it might be more convenient considering the administration intervals, thus allowing a higher adherence to therapy and patient autonomy [20].

Long-acting injectable Cabotegravir/Rilpivirine formulation

Cabotegravir (CR) is an integrase strand transfer inhibitor (INSTI) structurally similar to dolutegravir [48,49]. Both oral tablet or a long-acting (LA) injectable formulation are currently administered once daily in combination with rilpivirine [49,50]. The half-life of this drug is particularly long, ranging between 20 to 40 days [49]. CR is strongly bound to albumin and eliminated through the liver, even though no dose adjustments are required for patients with impaired hepatic function [49]. In turn, rilpivirine is an NNRTI with a long half-life too ranging from 30 to 90 days. It might also interfere with liver and pancreatic enzymes and cause headaches, nausea, dizziness, and fatigue [49].

Recently, the results from Phase III clinical trials (FLAIR and ATLAS) of this novel formulation, have been published [50]. FLAIR was an open-label Phase III trial in which the included patients were ART-naïve. Aside from K103N, none of them presented any other NNRTI resistance-associated mutations (RAMs) [49]. This trial was divided into two phases. The induction phase lasted for 20 weeks in which all patients took the oral fixed-dose combination of abacavir/ lamivudine/ dolutegravir. At 20th week, the maintenance phase began and all patients with less than 50 copies/ml were randomly distributed to a group that remained with the initial therapy and to another one that started a different oral therapy regimen with 30 mg cabotegravir + 25 mg rilpivirine once a day for 4 weeks. Following the 4 weeks, an intramuscular injection of 600 mg cabotegravir + 900 mg rilpivirine was administered, and afterward, a maintenance dose of LA cabotegravir 400 mg + 600 mg LA rilpivirine once a month [49,51,52]. After 48 weeks, the results showed that 7 patients out of 283 taking oral therapy (2.5%) had over 50 copies of viral RNA/ml as opposed to 6 patients out of 283 taking the LA formulation (2.1%). However, there were patients in both arms of the trial that withdrew from the study due to ineffectiveness of the treatment and few others confirmed the virologic failure in keeping viral loading under 200 copies/ml. Nevertheless, these results were enough to prove the non-inferiority of LA injectable therapy compared to oral therapy [46,49,51,53]. Patients were generally satisfied with this new therapy regimen and injection site reactions (ISRs) were mostly mild and constrained [49].

ATLAS was also an open-labeled and randomized trial, in which all the participants had less than 50 copies/ml of HIV RNA and followed an oral therapy regimen with 2 NRTI + 1 INSTI, NNRTI or PI for a minimum of 6 months. The patients were randomly distributed to continue with their usual regimen or take the LA therapy. Similarly, to the FLAIR study, the patients received 30 mg LA cabotegravir + 25mg LA rilpivirine for 4 weeks. Afterward, an intramuscular injection of 600 mg of LA cabotegravir + 900 mg LA rilpivirine was administered as a loading dose, and then, a maintenance dose of 400 mg LA cabotegravir + 600mg rilpivirine was taken once a month through the same route of administration [49,54,55]. The results showed that after 48 weeks only 5 out of 308 patients doing LA therapy had over 50 copies of viral RNA/ml (1.6%), compared to 1.0% (3/308) in the control group. This means that LA cabotegravir + rilpivirine therapy was considered non-inferior to the triple oral therapy. Contrary to the FLAIR study, there was a higher incidence of adverse reactions [46,49].

The LA injectable therapy has demonstrated significant advantages when compared to oral therapy in both studies.

Although most patients found this option preferable, there are still some concerns about this treatment option, such as the viral resistance, the compliance to this medication schedule, and how to proceed when an injection is missing [49]. Additionally, it is known that rifampicin activates the metabolism of CR, reducing its concentration in the plasma. This might cause a problem in HIV-positive patients co-infected with Tuberculosis since rifampicin is commonly used to treat it [49].

Despite these promising results, there are thus still some questions to be answered, including the effect of LA therapy on pregnant women. At the moment, there are other ongoing studies, for example, MOCHA that aims to provide information regarding the pharmacokinetics and safety of using LA CR on adolescents from 12 to 18 years old and ALTAS 2M in which a different regimen of 600 mg LA CR + 900 mg LA rilpivirine is taken every 8 weeks [49].

Fostemsavir

Fostemsavir was developed as a prodrug of temsavir, an attachment inhibitor. Its mechanism of action is innovative, as it connects to the viral glycoprotein gp120 which in turn is unable to bind to the CD4 receptor, and therefore, compromise the viral entry [48,56,57]. The BRIGHT study is an ongoing phase III clinical trial assessing the efficacy of this new drug. The included participants were infected with multi-resistant HIV-1 and with a viral loading over 400 copies/ml [57,58]. Patients were assigned into two cohorts: one randomized (RC) and another non-randomized (NRC). In the RC, patients took the placebo or 600 mg fostemsavir twice a day along with their failing ART drugs for 8 days. From the 9th day forward, all patients received 600 mg fostemsavir twice daily along with optimized background therapy just like the patients in the NRC [57,58]. After 24 weeks, 146 out of 272 patients in the RC (54%) had less than 40 copies of viral RNA/ml. In the NRC, the percentage of patients who had their viral loading less than 40 copies/ml was 36% (36/99) [57,58]. The results also showed a low prevalence of adverse effects, being headaches, nausea, and diarrhea the most frequent ones [57,58]. However, few patients experienced more severe adverse effects, such as acute renal failure and hepatocellular injury, among others [57]. Fostemsavir did not present major interactions with other classes of ARV drugs [57]. Nevertheless, it is still needed more information about the potential side effects and interactions of this new drug.

Leronlimab (PRO 140)

Leronlimab (PRO 140) is a humanized IgG4 monoclonal antibody. Its target is the CCR5 chemokine receptor since it binds to the extracellular domains of this molecule, i.e., the loop 2 and the N-terminus [59,60]. By this way, PRO 140 prevents virions from entering and infecting other host cells [59,60]. There are many similarities between Maraviroc and PRO 140 [59], and their difference in binding sites may suggest that these drugs could act by a synergistically way [61].

There are ongoing phase IIb/III trials aiming to evaluate the safety and effectiveness of leronlimab. In CD 02 trial, the efficacy of leronlimab has been tested in patients with an unsuccessful ART regimen. They took 350 mg of PRO 140 subcutaneously or placebo for 1 week along with their regular medication. One week later, participants took PRO 140 with an optimized regimen. Unfortunately, the results were not promising, since the failure rate was 76% due to the emergence of X4 viral strains for which PRO 140 was ineffective [59]. CD 03 is another clinical trial, in which participants are still being recruited. This trial intends to test the transition of stable patients treated with conventional therapy into

a weekly regimen of subcutaneous leronlimab in monotherapy, by assessing the viral loading for 48 weeks [62].

Overall, high tolerability and high threshold to the resistance of this new drug might allow it to be suitable for pre- and post-exposure prophylaxis [59].

UB-421

UB-421 is an Fc-glycosylated humanized Ig1 antibody that targets the CD4 receptor. It prevents the attachment and viral entry by attaching to domain 1 of that receptor and blocking it in a competitive manner [63].

The safety and efficacy of UB-421 have been assessed in several clinical trials. One of them is a phase II/III trial that started in September 2019 whereupon UB-421 was assessed along with a failing ART regimen at an initial stage for 1 week, and later, UB-421 was evaluated with an optimized background regimen for 24 weeks. The primary outcome measures were the viral loading log₁₀ difference from the baseline [64]. Additionally, a phase III study has just started in January 2020 with the main goals of evaluating the efficacy, safety, and tolerability of this drug as a monotherapy agent. Participants were distributed into two cohorts. Cohort 1 consisted of a control group taking standard c-ART, while in cohort 2 patients took only intravenous UB-421 at 25 mg/kg bi-weekly for 26 weeks. Afterward, all patients entered a follow-up phase receiving the standard c-ART. The primary outcome measure was the number of patients without virologic failure [65].

Vaccines

The eminent need for an effective vaccine against HIV has been quite discussed over several years. However, despite various attempts, this goal has not been achieved yet. To this date, there have been few effective clinical trials for vaccines against HIV. The most promising one was a phase III RV144 trial, but it showed a low level of protection against the virus [66,67] due to its resistance and difficulty to find a target that is common to many different strains [66,68].

Considering this extreme viral variability, a vaccine design based on live inactivated virus or attenuated virus might not be a viable strategy. Alternatively, the vaccines that have been currently tested, use attenuated viral proteins (e.g. gp120) to induce an immune response with the production of broadly neutralizing antibodies [69,70]. The target for neutralizing antibodies (nAb) is Env, and thus, this is a suitable immunogen to be considered when designing a vaccine. Another important aspect to consider is the delivery system for the vaccine. For example, nanoparticles were successfully tested as suitable carriers for vaccines [69,70].

The vesicular stomatitis virus has already been tested as a viral vector for expression of heterologous genes from other viruses, such as Ebola [71], and it is now being tested as a vector for the expression of the env glycoprotein of HIV [72]. Accordingly, an ongoing clinical trial is testing a therapeutic vaccine for HIV based on a multi-antigen DNA plasmid combined with an IL-12 plasmid (acting as adjuvants) and a vesicular stomatitis virus vector expressing the HIV-1 gag gene. The goal of this vaccine is to increase a humoral immune response in infected individuals and possibly reactivating memory CD4⁺ cells to eradicate the latent reservoirs [73].

A phase III clinical trial will be set to assess the efficacy of a new vaccine containing mosaic antigens, such as env, gag-pol, and gp140 (composed of gp120 plus the ectodomain of gp41) [74] obtained from an adenovirus serotype 26 vector [75]. This is an innovative challenge to formulate a “global vaccine” and overcome the major obstacle of the genetic diversity of HIV [76].

Novel Therapeutic Strategies

New Transdermal Drug Delivery Systems

Since most ARVs are administered orally, it is important to identify the main adversities of this route of administration, including those related to bioavailability, frequency of administration, and the hepatic first-pass metabolism. In particular, transdermal drug delivery systems (TDDS) are currently being investigated in order to diminish those issues [77-79]. The TDDS present an alternative to the conventional oral ARV regimens since this administration system allows obtaining a controlled and continuous drug release by avoiding the pharmacokinetic variations of the oral administration. This means that it may be possible to use drugs with a short half-life and find a simpler and acceptable regimen for patients [77,78,80].

Besides the reservoir and the drug itself (i.e. drug concentration, pka, molecular weight (ideally around 500 Da), log P (preferably ranged between 1 and 3), and the melting point) [77-79], a TDDS should have other essential components, as follows: a) permeation enhancers to increase the permeability through the stratum corneum (e.g. pyrrolidones, surfactants, phospholipids, and solvents); b) pressure-sensitive adhesives (PSA) to keep contact between the skin and the device; c) a release liner to cover the patch, and d) a backing layer which should not allow the diffusion of any excipients [77,78].

Transdermal delivery system for Tenofovir Alafenamide:

Tenofovir Alafenamide (TAF) is an NRTI commonly used for HIV treatment as part of oral therapy regimens. A controlled matrix delivery system was recently developed for this drug due to its low oral bioavailability [79]. In this system, the active substance is dispersed in a polymer matrix and a suitable solvent, which later evaporates, forming a drug reservoir. Then, the reservoir is shaped and interconnected with several layers. The overall system is composed of an adhesive layer that controls the release rate of the drug and it is in contact with the skin, followed by the drug reservoir and a second adhesive layer, connected to an exterior impermeable laminate [77-79]. Accordingly, this study [79] aimed to formulate a patch able to continuously release 8 mg/day TAF for one week. For that proposal, silicone-based or polyisobutylene (PIB) suspension patches and acrylate solution patches were used and several formulations were prepared varying the components, TAF concentration, and permeation enhancers for each patch. Specific parameters were determined, such as: a) crystallization of TAF in each matrix; b) effect of TAF particle homogenization; c) coat weight and TAF amount in each patch after exposure to stress conditions, and d) in vitro skin permeation studies using Franz diffusion cells. Overall, the suspension silicone patch formulated with 15% TAF (w/w), silicone, oleic acid, oleyl alcohol, and mineral oil as permeation enhancers, revealed to be the most suitable since it presented the target flux permeation rate of 7 µg/cm²/h through a 50 cm² patch area for an entire week, reaching a daily dose of 8.4 mg TAF. Additionally, this formulation was stable over time, non-irritating to the skin, and convenient when peeled off [79]. Despite these positive results and the promising chance of a new route of administration for TAF, further studies are still needed regarding both pharmacokinetic and safety of this new device [79].

Transdermal delivery of Enfuvirtide (T20) via ultrasounds:

As previously discussed, enfuvirtide (T20) is an entry inhibitor, usually administered by subcutaneous injection, 90 mg twice a day, which enables it rather inconvenient in terms of patient compliance. Thus, its transdermal delivery could be quite interesting option. However, there are some obstacles, including

the high molecular weight (4,492 Da) of this drug, which could compromise its diffusion through the stratum corneum [78,81]. Therefore, the ultrasound technique could be a suitable way to surpass this problem, since it reduces the barrier function of the stratum corneum [82,83]. Ultrasound waves are known to create pores that allow large molecules to cross the epidermis besides contributing to a fluid state of lipid skin layers, which in turn, promotes the transcellular pathway [83,84].

A recent study has assessed the effects of transdermal delivery of T20 using a low-frequency and low power ultrasound transducer patch in porcine models. The models were divided into 3 separate groups for 30 days as follows: one control group receiving injectable T20 twice a day; another group undergoing ultrasound treatment with saline solution and a third one treated with transdermal T20 via ultrasounds [81]. In this last case, T20 was in direct contact with the skin. The final device was obtained using wound dressing patches and a silicone ring that served as a reservoir, over which the ultrasound transducer was placed [81]. All groups were evaluated to understand the differences between them regarding skin health and bioavailability. The skin health criteria were based on histologic cuts and trans-epidermal water loss. The bioavailability of T20 was evaluated through liquid chromatography/electrospray ionization mass spectrometry (LC-MS/MS) [81]. Overall, no significant differences between the saline group and the transdermal T20 group were observed, which indicates that ultrasound did not affect the skin. The histologic cuts showed mild signs of inflammation in the active patch group. In addition, the animals from the active patch group had a longer Tmax and a lower Cmax when compared to the injectable T20 group. The plasma concentrations were generally lower with the transdermal treatment as expected [81].

Nanosystems for Drug Delivery

Nanotechnology has contributed to several applications in drug delivery through different routes of administration and overcoming certain formulation obstacles, such as solubility, bioavailability, and drug stability [85]. Additionally, nanosystems could be a promising strategy for targeted therapy since they allow the encapsulation of drugs or specific genes that could be transported not only to infected cells but also to reservoir tissues, including the central nervous system and lymph nodes, thereby potentially eradicating the virus [85].

There are several types of nanosystems that could function as carriers for ARV drugs, such as liposomes, niosomes, solid-lipid or polymeric (e.g. PGLA) or diamond nanoparticles, dendrimers, among others (Figure 1) [85]. In general, liposomes and niosomes are less toxic and cost-effective. Both are made of an aqueous compartment surrounded by a lipid bilayer and could be a promising alternative for drug delivery as these carriers can be easily absorbed by macrophages [86]. For example, stavudine was already encapsulated into gelatin nanoparticles, which in turn were incorporated into liposomes. Accordingly, several formulations were successfully prepared in this report and characterized in terms of drug release, cytotoxicity, and hemocompatibility [86].

Dendrimers are somewhat toxic but have numerous reactive groups capable of forming conjugates for targeted drug delivery [85,87]. Dendrimers possess a typical three-dimensional branched structure where the outer layers are appropriate for conjugation, while the inner layers are quite effective for drug encapsulation, resulting in a controlled drug release [87]. In a recent study published in 2013, zidovudine was encapsulated into poly (propyl ether imine) dendrimers in an attempt to surpass the short half-life and provide a more continuous release of this drug [88].

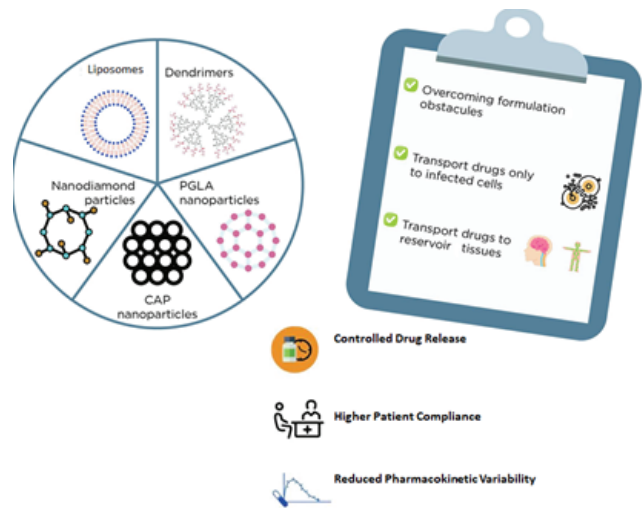


Figure 1. Examples nanosystems for drug delivery and the main advantages of their usage.

The study showed that not only this delivery system reduced the hemolytic effect, but it also prolonged the drug release decreasing the occurrence of side effects [88]. In another study, carbosilane dendrimers (G3-S16 and G2-NF16) were used to encapsulate zidovudine, efavirenz, and tenofovir [89]. The encapsulated drugs were tested for antiviral activity in PBMC cells and TMZ-bl cells infected with X4 and R5 viral strains. The results showed an enlarged antiviral activity of all three drugs when formulated with dendrimers [89].

Polymeric nanoparticles, such as chitosan and poly (lactide co-glycolide), are known to be very effective in drug delivery while exhibiting low toxicity similar to metal gold or silver nanoparticles [85]. In particular, nanoparticles have been tested to improve ARV delivery to the central nervous system since many ARV drugs suffer efflux mechanisms, which may contribute to the spreading of this virus, and thereby, to the development of HIV-related neurologic disorders [90,91]. For example, transferrin is one of the nanoparticles studied for this purpose due to the abundance of transferrin receptors in the blood-brain barrier [85,90]. In a recent study, nanodiamond particles were studied in order to load efavirenz (EFV) and deliver it to the brain [92]. This study compared nanodiamond particles (ND) with both unmodified and modified surface (ND-COOH and ND-NH₂) in terms of toxicity, and drug loading capacity. ND-COOH was found to be less suitable than the other two since it induced a higher production of reactive oxygen species (ROS) [92]. On the contrary, the formulation with unmodified nanodiamond particles (ND-EFV) presented a suitable and slower release profile through a blood-brain barrier model and it was able to impair viral replication for a longer period in comparison with free EFV [92]. Overall, this study suggests that ND particles are a promising drug delivery system, due to their nontoxic nature and ability to cross the blood-brain barrier. However, further studies still need to be performed to evaluate the effect in in vivo models and possible side effects, since ND particles may interfere with the expression of genes related to neuronal function [92]. Another recent study attempted to demonstrate the effects of using PGLA nanoparticles loaded with EFV and saquinavir (SQV) [93]. In general, ARV formulated with PGLA nanoparticles showed lower IC₅₀ values in comparison with the free ARV drugs. The release profile of these nanoparticles was also quite favorable, since the drugs were first

Table 3. Examples of main outcomes from in vitro studies of ARV loaded nanocarriers for HIV management.

Nanocarriers + ARV	Main Outcomes	References
Liposomes + Stavudine	Liposomes were revealed to be a promising alternative for stavudine delivery as these carriers can be easily absorbed by macrophages.	[87]
Dendrimer + Zidovudine	The formulation reduced the AZT hemolytic effect and prolonged the drug release, decreasing the occurrence of side effects.	[89]
Carbosilane Dendrimers + Zidovudine Carbosilane Dendrimers + Efavirenz Carbosilane Dendrimers + Tenofovir	An enlarged antiviral activity of all three drugs was observed when formulated with dendrimers.	[90]
Nanodiamond Particles + Efavirenz	A suitable and slower release profile through a blood-brain barrier model was obtained impairing viral replication for a longer period.	[93]
PGLA nanoparticles + Efavirenz PGLA nanoparticles + Saquinavir	An enlarged antiviral activity of all three drugs was obtained with PGLA nanoparticles.	[94]
PGLA nanoparticles + Efavirenz + Raltegravir (thermosensitive gel)	A lower EC90 and a constant release of these loaded drugs were obtained being a promising option for pre-exposure HIV prophylaxis.	[95]
CAP nanoparticles + Efavirenz (thermosensitive gel)	High encapsulation efficacy and lower cytotoxicity in HeLa cells were observed besides enhanced prophylactic activity in TMZ-bl cells treated with EFV-CAP nanoparticles.	[97]
CAP nanoparticles + Dolutegravir (thermosensitive gel)	pH (4.2 and 7.4) influenced both the drug release and the cytotoxicity of this formulation.	[96]

rapidly released, and then, at a continuous rate. Additionally, when adding free tenofovir to these nanoparticles, a synergic effect was obtained resulting in dose reduction to impair HIV activity [93]. In another report, PGLA nanoparticles were loaded with raltegravir (RAL) and EFV, and further incorporated in a thermo-sensitive vaginal gel for HIV prophylaxis [94]. The goal was to prepare a gel that would acquire a gel-like texture at 37°C and liquid at room temperature. The PGLA nanoparticles were here prepared via a modified emulsion-solvent evaporation method, and characterized through multiple experiments, including in vitro release studies with human cervical cells. When compared with an EFV-RAL solution, the loaded nanoparticles had a lower EC90 and were able to ensure a constant release of these drugs, despite their different intracellular concentrations and routes of metabolism [94]. Overall, this formulation was considered a successful and promising option for pre-exposure HIV prophylaxis [94]. Still regarding prophylaxis, cellulose acetate-phthalate (CAP) nanoparticles have shown promising outcomes when incorporated into thermosensitive gels. Although most commonly used as a coating agent for other formulations, CAP was found to possess ARV activity by promoting viral disintegration and interfering with the mechanisms of viral entry. Moreover, CAP was stable at low pH, which facilitated a vaginal drug delivery [95,96]. In another study [96], a thermosensitive gel was formulated using EFV-loaded CAP nanoparticles. This formulation was assessed for cytotoxicity and prophylactic activity in human cervical cells (HeLa) and TZM-bl cells against an EFV solution. The results revealed a remarkable encapsulation efficacy as well as lower cytotoxicity in HeLa cells treated with EFV-CAP nanoparticles formulated in the thermosensitive gel. Moreover, these nanoparticles showed enhanced prophylactic activity (at 5 ng/ml) in TMZ-bl cells when compared with the EFV solution [96].

Similarly, dolutegravir (DTG) loaded CAP nanoparticles were incorporated into a thermosensitive gel and tested at pH= 4.2 and pH=7.4 to simulate vaginal and seminal fluid conditions, respectively [95]. In this study, the pH clearly influenced both the

drug release and the cytotoxicity of the tested formulation [95].

In summary, the examples of the most recent in vitro studies of ARV loaded nanocarriers for HIV management are listed in (Table 3).

Conclusion and Future Perspectives

HIV infection is still considered one of the major pandemics worldwide. In the 90s, with the emergence of the first antiretroviral drugs and c-ART, the status of this infection changed from deadly illness to chronic infection, allowing infected people to live as long as non-infected individuals. However, c-ART presents many downsides, such as side effects, frequency of drug administration, and the possibility of viral resistance, while it still not provides a definitive cure.

Several studies have been made to shift the therapeutic strategies towards this infection. Accordingly, new drugs from the already well-known ARV drug classes and others with a novel therapeutic target have been evaluated in clinical trials. Moreover, some classical ARV drugs have been also studied with a different route of administration and/ or formulation, enabling a prolonged drug release as well as surpassing some compliance issues derived from polymedication in HIV patients. In fact, nanotechnology has been shown to provide a more targeted and controlled release of antiretrovirals overcoming several formulation obstacles including cytotoxicity. Nevertheless, further investigation is urgently needed especially regarding prophylactic vaccines and pharmacokinetic/ pharmacodynamic in silico prediction followed by well-defined in vivo studies.

Funding

This work was supported with Gilead GENESE Ref. PGG/006/2016

References

1. Maartens G, Celum C, Lewin SR. HIV infection: epidemiology, pathogenesis, treatment, and prevention. *Lancet*. 2014; 384: 258-271.

2. UNAIDS. Global HIV & AIDS statistics — 2019 fact sheet | UNAIDS [Internet]. [cited 2019 Nov 10]. Available from: <https://www.unaids.org/en/resources/fact-sheet>
3. German Advisory Committee Blood (Arbeitskreis Blut), Subgroup 'Assessment of Pathogens Transmissible by Blood' GACB (Arbeitskreis, Blood) S 'Assessment of PT by Human Immunodeficiency Virus (HIV). *Transfus Med Hemother*. 2016; 43: 203-222.
4. Wilen CB, Tilton JC, Doms RW. HIV: cell binding and entry. *Cold Spring Harb Perspect Med*. 2012; 2: a006866.
5. Casadellà M, Cozzi-Lepri A, Phillips A, Noguera-Julian M, Bickel M, et al. Plasma HIV-1 Tropism and the Risk of Short-Term Clinical Progression to AIDS or Death. *PLoS One*. 2017; 12: e0166613.
6. Sierra S, Kupfer B, Kaiser R. Basics of the virology of HIV-1 and its replication. *J Clin Virol*. 2005; 34: 233-244.
7. Krogstad P. Molecular Biology of the Human Immunodeficiency Virus: Current and Future Targets for Intervention. *Semin Pediatr Infect Dis*. 2003; 14: 258-268.
8. Engelman A, Cherepanov P. The structural biology of HIV-1: mechanistic and therapeutic insights. *Nat Rev Microbiol*. 2012; 10: 279-290.
9. Shaw GM, Hunter E. HIV transmission. *Cold Spring Harb Perspect Med*. 2012; 2: a006965.
10. Cohen MS, Shaw GM, McMichael AJ, Haynes BF. Acute HIV-1 Infection. *N Engl J Med*. 2011; 364: 1943-1954.
11. Fanales-Belasio E, Raimundo M, Suligoi B, Buttò S. HIV virology and pathogenetic mechanisms of infection: A brief overview. *Ann Ist Super Sanita*. 2010; 46: 5-14.
12. Zulfiqar HF, Javed A, Sumbal, Afroze B, Ali Q, et al. HIV Diagnosis and Treatment through Advanced Technologies. *Front Public Heal*. 2017; 5: 32.
13. Kemnic TR, Gulick PG. HIV Antiretroviral Therapy. StatPearls. StatPearls Publishing; 2019.
14. Desai M, Iyer G, Dikshit RK. Antiretroviral drugs: critical issues and recent advances. *Indian J Pharmacol*. 2012; 44: 288-298.
15. Arts EJ, Hazuda DJ. HIV-1 Antiretroviral Drug Therapy. *Cold Spring Harb Perspect Med*. 2012; 2: a007161.
16. Lange JMA, Ananworanich J. The discovery and development of antiretroviral agents. *Antivir Ther*. 2014; 19:5-14.
17. Weller I V, Williams IG. ABC of AIDS. Antiretroviral drugs. *BMJ*. 2001; 322: 1410-1412.
18. Van Der Ryst E. Maraviroc - A CCR5 Antagonist for the Treatment of HIV-1 Infection. *Front Immunol*. 2015; 6: 277.
19. CHMP. Celsentri - EPAR summary for the public. 2007.
20. Bruno CJ, Jacobson JM. Ibalizumab: an anti-CD4 monoclonal antibody for the treatment of HIV-1 infection. *J Antimicrob Chemother*. 2010; 65: 1839-1841.
21. Tseng A, Hughes CA, Wu J, Seet J, Phillips EJ. Cobicistat Versus Ritonavir: Similar Pharmacokinetic Enhancers But Some Important Differences. *Ann Pharmacother*. 2017; 51: 1008-1022.
22. von Hentig N. Clinical use of cobicistat as a pharmacoenhancer of human immunodeficiency virus therapy. *HIV AIDS (Auckl)*. 2016; 8: 1-16.
23. Ashkenazi A, Wexler-Cohen Y, Shai Y. Multifaceted action of Fuzeon as virus-cell membrane fusion inhibitor. *Biochim Biophys Acta - Biomembr*. 2011; 1808: 2352-2358.
24. Ding X, Zhang X, Chong H, Zhu Y, Wei H, et al. Enfuvirtide (T20)-Based Lipopeptide Is a Potent HIV-1 Cell Fusion Inhibitor: Implications for Viral Entry and Inhibition. *J Virol*. 2017; 91: e00831-17.
25. Poveda E, Briz V, Soriano V. Enfuvirtide, the first fusion inhibitor to treat HIV infection. *AIDS Rev*. 7: 139-147.
26. Kitchen CM, Nuño M, Kitchen SG, Krogstad P. Enfuvirtide antiretroviral therapy in HIV-1 infection. *Ther Clin Risk Manag*. 2008; 4: 433-439.
27. AIDSinfo. FDA Approval of HIV Medicines | AIDSinfo [Internet]. [cited 2019 Sep 17]. Available from: <https://aidsinfo.nih.gov/understanding-hiv-aids/infographics/25/fda-approval-of-hiv-medicines>
28. CHMP. Prezista | European Medicines Agency [Internet]. Available from: <https://www.ema.europa.eu/en/medicines/human/EPAR/prezista>
29. Agency EM. Vitekta: Withdrawal of the marketing authorisation in the European Union [Internet]. Available from: https://www.ema.europa.eu/en/documents/public-statement/public-statement-vitekta-withdrawal-marketing-authorisation-european-union_en.pdf
30. HIV/AIDS Historical Time Line 1995-1999 | FDA [Internet]. [cited 2019 Sep 17]. Available from: <https://www.fda.gov/patients/hiv-timeline-and-history-approvals/hivaids-historical-time-line-1995-1999>
31. Zhu Y, Zhang X, Ding X, Chong H, Cui S, et al. Exceptional potency and structural basis of a T1249-derived lipopeptide fusion inhibitor against HIV-1, HIV-2, and simian immunodeficiency virus. *J Biol Chem*. 2018; 293: 5223-5334.
32. Pu J, Wang Q, Xu W, Lu L, Jiang S. Development of protein-and peptide-based hiv entry inhibitors targeting gp120 or gp41. Vol. 11, Viruses. MDPI AG; 2019.
33. Chong H, Xue J, Zhu Y, Cong Z, Chen T, et al. Monotherapy with a low-dose lipopeptide HIV fusion inhibitor maintains long-term viral suppression in rhesus macaques. Silvestri G, editor. *PLoS Pathog*. 2019; 15: e1007552.
34. Woollard SM, Kanmogne GD. Maraviroc: a review of its use in HIV infection and beyond. *Drug Des Devel Ther*. 2015; 9: 5447-5468.
35. CHMP. Genvoya, INN-Elvitegravir/Cobicistat/Emtricitabine/Tenofovir Alafenamide (as fumarate). 2015.
36. CHMP. Atripla, INN-Efavirenz/Emtricitabine/Tenofovir disoproxil (as fumarate) - ANNEX I SUMMARY OF PRODUCT CHARACTERISTICS. 2007.
37. CHMP. Rezolsta, INN-darunavir,cobicistat - ANNEX I SUMMARY OF PRODUCT CHARACTERISTICSANNEX I SUMMARY OF PRODUCT CHARACTERISTICS. 2014.
38. CHMP. Triumeq, INN-dolutegravir, abacavir, lamivudine - ANNEX I SUMMARY OF PRODUCT CHARACTERISTICS. 2014.
39. CHMP. Evotaz, INN - atazavir,cobicistat - ANNEX I SUMMARY OF PRODUCT CHARACTERISTICS. 2015.
40. CHMP. Descovy, INN-Emtricitabine/Tenofovir Alafenamide - ANNEX I SUMMARY OF PRODUCT CHARACTERISTICS. 2016.
41. Xu W, Li H, Wang Q, Hua C, Zhang H, et al. Advancements in Developing Strategies for Sterilizing and Functional HIV Cures. *Biomed Res Int*. 2017; 2017: 6096134.
42. Kumar R, Qureshi H, Deshpande S, Bhattacharya J. Broadly neutralizing antibodies in HIV-1 treatment and prevention. *Ther Adv vaccines Immunother*. 2018; 6: 61-68.
43. Awi NJ, Teow S-Y. Antibody-Mediated Therapy against HIV/AIDS: Where Are We Standing Now? *J Pathog*. 2018 3; 2018: 1-9.
44. Markham A. Ibalizumab: First Global Approval. *Drugs*. 2018; 78: 781-785.
45. Agency EM. Trogarzo: Pending EC decision | European Medicines Agency [Internet]. [cited 2019 Jul 29]. Available from: <https://www.ema.europa.eu/en/medicines/human/summaries-opinion/trogarzo>

46. Singh K, Sarafianos SG, Sönnnerborg A. Long-Acting Anti-HIV Drugs Targeting HIV-1 Reverse Transcriptase and Integrase. *Pharmaceuticals (Basel)*. 2019; 12: 62.
47. Iacob SA, Iacob DG. Ibalizumab Targeting CD4 Receptors, An Emerging Molecule in HIV Therapy. *Front Microbiol*. 2017; 8: 2323.
48. Zhang X. Anti-retroviral drugs: current state and development in the next decade. *Acta Pharm Sin B*. 2018; 8: 131-136.
49. Fernandez C, van Halsema CL. Evaluating cabotegravir/rilpivirine long-acting, injectable in the treatment of HIV infection: emerging data and therapeutic potential. *HIV AIDS (Auckl)*. 2019; 11: 179-192.
50. Collins S. (HIV i-B. HIV pipeline 2019 report. 2019; 8591: 1-10.
51. Chloe Orkin, Keikawus Arastéh, Miguel Górgolas Hernández-Mora, Vadim Pokrovsky4, Edgar T. Overton, Pierre-Marie Girard, Shinichi Oka, Ronald D'Amico, David Dorey, Sandy Griffith, David A. Margolis, Peter E. Williams, Wim Parys WS. LONG-ACTING CABOTEGRAVIR + RILPIVIRINE FOR HIV MAINTENANCE: FLAIR WEEK 48 RESULTS | CROI Conference [Internet]. Conference on Retroviruses and Opportunistic Infections Seattle, Washington. Available from: <http://www.croiconference.org/sessions/long-acting-cabotegravir-rilpivirine-hiv-maintenance-flair-week-48-results>
52. ViiV Healthcare. Study to Evaluate the Efficacy, Safety, and Tolerability of Long-acting Intramuscular Cabotegravir and Rilpivirine for Maintenance of Virologic Suppression Following Switch From an Integrase Inhibitor in HIV-1 Infected Therapy Naive Participants [Internet]. ClinicalTrials.gov. [cited 2019 Oct 18]. Available from: <https://clinicaltrials.gov/ct2/show/NCT02938520>
53. AIDSinfo. Virologic Failure | Definition | AIDSinfo [Internet]. Available from: <https://aidsinfo.nih.gov/understanding-hiv-aids/glossary/879/virologic-failure>
54. Simon Collins. Phase 3 results with dual therapy cabotegravir/rilpivirine long-acting injections: ATLAS and FLAIR studies | HTB | HIV i-Base [Internet]. HIV i-Base. 2019 [cited 2019 Sep 26]. Available from: <http://i-base.info/htb/35812>
55. ViiV Healthcare. Study Evaluating the Efficacy, Safety, and Tolerability of Switching to Long-acting Cabotegravir Plus Long-acting Rilpivirine From Current Antiretroviral Regimen in Virologically Suppressed HIV-1-infected Adults - Full Text View - ClinicalTrials.gov [Internet]. ClinicalTrials.gov. 2016 [cited 2019 Oct 18]. Available from: <https://clinicaltrials.gov/ct2/show/NCT02951052>
56. Gulick RM. Investigational Antiretroviral Drugs: What is Coming Down the Pipeline. *Top Antivir Med*. 2018; 25: 127-132.
57. Cahn P, Fink V, Patterson P. Fostemsavir: a new CD4 attachment inhibitor. *Curr Opin HIV AIDS*. 2018; 13: 341-345.
58. Proudfoot C, Ackerman P, Llamoso C, Cella D, Clark A, et al. 547. Results of Patient-Reported Outcome Data From the Phase III BRIGHT Study of Fostemsavir. In: Open Forum Infectious Diseases. Oxford University Press; 2018; S203-S203.
59. Thompson MA. The return of PRO 140, a CCR5-directed mAb. *Curr Opin HIV AIDS*. 2018; 13: 346-353.
60. Dhody K, Pourhassan N, Kazempour K, Green D, Badri S, et al. PRO 140, a monoclonal antibody targeting CCR5, as a long-acting, single-agent maintenance therapy for HIV-1 infection. *HIV Clin Trials*. 2018; 19: 85-93.
61. Khatib N, Das S. PRO 140 - A Novel CCR5 Co-Receptor Inhibitor. *Recent Pat Antiinfect Drug Discov*. 2009; 5: 18-22.
62. CytoDyn I. Study of PRO 140 SC as Single Agent Maintenance Therapy in Virologically Suppressed Subjects With CCR5-tropic HIV-1 Infection - Full Text View - ClinicalTrials.gov [Internet]. ClinicalTrials.gov. [cited 2019 Oct 18]. Available from: <https://clinicaltrials.gov/ct2/show/NCT02859961>
63. Wang CY, Wong WW, Tsai HC, Chen YH, Kuo BS, et al. Effect of anti-CD4 antibody UB-421 on HIV-1 rebound after treatment interruption. *N Engl J Med*. 2019; 380: 1535-1545.
64. United BioPharma. A Multicenter, Single-Arm, 24-Week Study of UB-421 in Combination With Optimized Background Therapy (OBT) Regimen in Patients With Multi-Drug Resistant (MDR) HIV-1 Infection [Internet]. ClinicalTrials.gov. 2017. Available from: <https://clinicaltrials.gov/ct2/show/study/NCT03164447>
65. United BioPharma. A Phase III, Randomized, Open-label, Controlled Trial to Investigate the Efficacy and Safety of UB-421 Monotherapy as Substitution for Stable Antiretroviral Therapy in HIV-1 Infected Adults [Internet]. ClinicalTrials.gov. 2017. Available from: <https://clinicaltrials.gov/ct2/show/study/NCT03149211>
66. Brett-Major DM, Crowell TA, Michael NL. Prospecting for an HIV vaccine. *Trop Dis Travel Med Vaccines*. 2017; 3.
67. Lema D, Garcia A, De Sanctis JB. HIV vaccines: A brief overview. *Scand J Immunol*. 2014; 80: 1-11.
68. Robinson HL. HIV/AIDS Vaccines: 2018. *Clin Pharmacol Ther*. 2018; 104: 1062-1073.
69. Aikins ME, Bazzill J, Moon JJ. Vaccine nanoparticles for protection against HIV infection. *Nanomedicine*. 2017; 12: 673-682.
70. Brinkkemper M, Slieden K. Nanoparticle Vaccines for Inducing HIV-1 Neutralizing Antibodies. *Vaccines*. 2019; 7: 76.
71. Clarke DK, Hendry RM, Singh V, Rose JK, Seligman SJ, et al. Live virus vaccines based on a vesicular stomatitis virus (VSV) backbone: Standardized template with key considerations for a risk/benefit assessment. *Vaccine*. 2016; 34: 6597-6609.
72. Racine T, Kobinger GP, Arts EJ. Development of an HIV vaccine using a vesicular stomatitis virus vector expressing designer HIV-1 envelope glycoproteins to enhance humoral responses. *AIDS Res Ther*. 2017; 14: 55.
73. National Institute of Allergy and Infectious Diseases (NIAID); Profectus BioSciences INI of HCC (CC). Therapeutic Vaccine for HIV - Full Text View - ClinicalTrials.gov [Internet]. [cited 2019 Nov 10]. Available from: <https://clinicaltrials.gov/ct2/show/NCT01859325>
74. Harris A, Borgnia MJ, Shi D, Bartesaghi A, He H, et al. Trimeric HIV-1 glycoprotein gp140 immunogens and native HIV-1 envelope glycoproteins display the same closed and open quaternary molecular architectures. *Proc Natl Acad Sci U S A*. 2011; 108: 11440-11445.
75. Janssen Vaccines & Prevention B.V. A Study of Heterologous Vaccine Regimen of Adenovirus Serotype 26 Mosaic4 Human Immunodeficiency Virus(Ad26.Mos4.HIV), Adjuvanted Clade C gp140 and Mosaic gp140 to Prevent HIV-1 Infection Among Cis-gender Men and Transgender Individuals Who Have Sex With [Internet]. ClinicalTrials.gov. 2019 [cited 2019 Nov 13]. Available from: <https://clinicaltrials.gov/ct2/show/NCT03964415>
76. Mega ER. 'Mosaic' HIV vaccine to be tested in thousands of people across the world. *Nature*. 2019; 572: 165-166.
77. Singh N, Singh R, Verma V. Advances in Biology & BioMedicine An Introduction to the Transdermal Delivery of Antiretrovirals. 2014;
78. Ham AS, Buckheit RW Jr. Current and emerging formulation strategies for the effective transdermal delivery of HIV inhibitors. *Ther Deliv*. 2015; 6: 217-229.
79. Puri A, Bhattacharjee SA, Zhang W, Clark M, Singh O, et al. Development of a Transdermal Delivery System for Tenofovir Alafenamide, a Prodrug of Tenofovir with Potent Antiviral Activity Against HIV and HBV. *Pharmaceuticals*. 2019; 11: 173.
80. Vedha Hari BN, Devendharan K, Narayanan N. Approaches of novel drug delivery systems for Anti-HIV agents. *Int J Drug Dev Res*. 2013; 5: 16-24.
81. Snook KA, Van Ess R, Werner JR, Clement RS, Ocon-Grove OM, et al. Transdermal Delivery of Enfuvirtide in a Porcine Model

- Using a Low-Frequency, Low-Power Ultrasound Transducer Patch. *Ultrasound Med Biol.* 2019; 45: 513-525.
82. Tachibana K, Tachibana S. The use of ultrasound for drug delivery. *Echocardiography.* 2001; 18: 323-328.
83. Luis J, Park EJ, Meyer RJ, Smith NB. Rectangular cymbal arrays for improved ultrasonic transdermal insulin delivery. *J Acoust Soc Am.* 2007; 122: 2022-2030.
84. Marwah H, Garg T, Goyal AK, Rath G. Permeation enhancer strategies in transdermal drug delivery. *Drug Deliv.* 2016; 23: 564-578.
85. Grande F, Ioele G, Occhiuzzi MA, De Luca M, Mazzotta E, et al. Reverse transcriptase inhibitors nanosystems designed for drug stability and controlled delivery. *Pharmaceutics.* 2019; 11: 197.
86. Nayak D, Boxi A, Ashe S, Thathapudi NC, Nayak B. Stavudine loaded gelatin liposomes for HIV therapy: Preparation, characterization and in vitro cytotoxic evaluation. *Mater Sci Eng C.* 2017; 73 :406-416.
87. Mhlwatika Z, Aderibigbe BA. Application of dendrimers for the treatment of infectious diseases. *Molecules.* 2018; 23: 2205.
88. Kumar S. In-vitro and in-vivo Evaluation of Poly (Propyl Ether Imine) (PETIM) Dendrimer for Sustained Delivery of Zidovudine. *J Antivir Antiretrovir.* 2013; 05.
89. Vacas-Córdoba E, Galán M, de la Mata FJ, Gómez R, Pion M, et al. Enhanced activity of carbosilane dendrimers against HIV when combined with reverse transcriptase inhibitor drugs: Searching for more potent microbicides. *Int J Nanomedicine.* 2014; 9: 3591-3600.
90. Gomes MJ, Neves J das, Sarmiento B. Nanoparticle-based drug delivery to improve the efficacy of antiretroviral therapy in the central nervous system. *Int J Nanomedicine.* 2014; 9: 1757-1769.
91. Curley P, Liptrott NJ, Owen A. Advances in nanomedicine drug delivery applications for HIV therapy. Vol. 4, Future Science OA. Future Medicine Ltd.; 2018.
92. Roy U, Drozd V, Durygin A, Rodriguez J, Barber P, et al. Characterization of Nanodiamond-based anti-HIV drug Delivery to the Brain. *Sci Rep.* 2018; 8.
93. Chaowanachan T, Krogstad E, Ball C, Woodrow KA. Drug synergy of tenofovir and nanoparticle-based antiretrovirals for HIV prophylaxis. *PLoS One.* 2013; 8: e61416.
94. Date AA, Shibata A, Goede M, Sanford B, La Bruzzo K, et al. Development and evaluation of a thermosensitive vaginal gel containing raltegravir+efavirenz loaded nanoparticles for HIV prophylaxis. *Antiviral Res.* 2012; 96: 430-436.
95. Mandal S, Khandalavala K, Pham R, Bruck P, Varghese M, et al. Cellulose Acetate Phthalate and Antiretroviral Nanoparticle Fabrications for HIV Pre-Exposure Prophylaxis. *Polymers (Basel).* 2017; 9: 423.
96. Date AA, Shibata A, McMullen E, La Bruzzo K, Bruck P, et al. Thermosensitive Gel Containing Cellulose Acetate Phthalate-Efavirenz Combination Nanoparticles for Prevention of HIV-1 Infection. *J Biomed Nanotechnol.* 2015; 11: 416-427.

***Correspondence:** Andreia Ascenso, Department of Pharmaceutical Technology, Portugal, E-mail: andreaascenso@ul.pt / r.cunha@campus.ul.pt

Rec: 02 Apr 2021; **Acc:** 19 Apr 2021; **Pub:** 23 Apr 2021

J Clin Case Rep Rev. 2021;4(2):166
DOI: 10.36879/JCCRR.21.000166

Copyright © 2021 The Author(s). This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International License (CCBY).